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Hygrophiloside tetra-acetate (1a). Prepared by acetylation of 1 (Ac₂O, pyridine, room temp., 2 hr). Crystallization from EtOH gave pure 1a, mp 148–148.5°; $[\alpha]_D^{20}-69^\circ$ (CHCl₃; c 0.9); ¹H NMR (90 MHz, CDCl₃): δ9.60 (s, CHO-10), 6.78 (m, H-7), 6.23 (d, J=2 Hz, H-1), 6.12 (d, J=6.5 Hz, H-3), 4.92 (d, J=6.5 Hz, H-4), 3.39 (m, H-9), 3.12 (s, OH), 2.91 and 2.68 (br ABsystem, J=19 Hz, CH₂-6), 2.11, 2.03, 2.00 and 1.98 (4 × OAc); ¹³C NMR (22.6 MHz, CDCl₃): δ189.3 (C-10), 152.3 (C-7), 142.9 (C-8), 139.4 (C-3), 110.0 (C-4), 91.4 (C-1),73.1 (C-5), 52.7 (C-9), 46.0 (C-6), 96.0, 71.0, 71.8, 68.2, 71.8, 61.5 (C-1–C-6 in the β-glucopyranosyl moiety). (Found: C, 52.7; H, 5.5. C₂₃H₂₈O₁₃· 1/2 H₂O requires: C, 53.0; H, 5.6%)

Conversion to isoaucubin penta-acetate (2a). To a stirred soln of 1 (90 mg) in MeOH (5 ml) was added NaBH₄ (60 mg). After 45 min the mixture was taken to dryness and the residue acetylated as above. Work-up gave, after prep. TLC, 2a (78 mg) as crystals from EtOH, mp and mmp 121-122°; $\begin{bmatrix} \alpha \end{bmatrix}_{0}^{20} - 91^{\circ}$ (CHCl₃; c 0.7), ltt. [5] mp 125°; $\begin{bmatrix} \alpha \end{bmatrix}_{0}^{18} - 46^{\circ}$ (EtOH; c 0.95); an authentic sample, kindly supplied by Dr. Endo, exhibited the

rotation $[\alpha]_D^{20} - 90^\circ$ (CHCl₃; c 0.2). The ¹H and ¹³C NMR spectra were as reported [5].

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A NOR-SESQUITERPENE-γ-LACTONE FOUND IN CREPIS PYGMAEA

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Key Word Index—*Crepis pygmaea*; Compositae; sesquiterpenoid lactones; 1,2–4,5-tetrahydro-11-nor-11-hydroxy-Δ^{7,11}-santonin.

Abstract—The isolation and structural elucidation of a novel nor-sesquiterpene-y-lactone are reported.

INTRODUCTION

We have recently reported the isolation and structural elucidation of an unusual nor-sesquiterpene-γ-lactone (1) found in Crepis pygmaea L.‡[1], whose structure has been definitively confirmed through synthesis [2]. During the isolation of 1 from the chloroform extract of the whole plant, a number of minor by-products were observed.

The present report describes the characterization of one of these products as the novel nor-sesquiterpenoid, 1,2-4,5-tetrahydro-11-nor-11-hydroxy- $\Delta^{7,11}$ -santonin (2) (or its mirror image).

‡Plant material was collected in July-August on Vettore mountain, Umbria, Italy. A specimen (voucher Nos. 2115/01) has been deposited at The University of Perugia, Perugia, Italy.

RESULTS AND DISCUSSION

Compound 2 was obtained by chromatographic fractionation of the chloroform extract of *C. pygmaea* as previously described [1]. In particular, repeated fractionation on silica gel (Kieselgel 60 Merck) columns of the crude fraction containing 1 [1] yielded, besides 1, chromatographically pure 2 (80 mg), which crystallized from ether as white cubes.

Compound 2, mp 177–179°; R_f (TLC) 0.39 (pre-coated Merck plates, n-hexane—CH₂Cl₂-i-PrOH, 34:6:10), $[\alpha]_D$ + 25.7° (CHCl₃; c 2.9); MS m/z 250 [M]⁺; UV λ_{EOH}^{EOH} nm (ϵ): 237 (8420); molecular formula C₁₄H₁₈O₄ (Found: C, 66.92; H, 7.32. C₁₄H₁₈O₄ requires: C, 67.18; H, 7.25%). The IR spectrum ($\nu_{Max}^{CHCl_3}$ cm⁻¹: 3520, 1755 and 1710) suggested the presence of an α,β -unsaturated γ -lactone containing a cyclohexanone ring and an additional alcoholic function. The ¹H NMR spectrum

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Table 1. ¹H NMR spectral data of compound 2 (500 MHz, CHCl₃, TMS as internal standard)

Н	δ	Н	δ
1 ax	1.57 ddd	8 ax	2.35 dddd
1 eq	1.77 ddd	8 eq	2.89 ddd
2 eq	2.40 ddd	9 ax	1.28 ddd
2 ax	2.54 ddd	9 eq	1.71 ddd
4	2.60 ddq	14	1.22 d
5	1.27 dd	15	1.24 s
6	4.71 ddd		

J (Hz): 1ax, 1eq = 14.0; 1ax, 2eq = 5.2; 1ax, 2ax = 14.0; 1eq, 2ax = 6.6; 1eq, 2eq = 2.2; 2ax, 2eq = 14.0; 4, 5 = 11.6; 4, 6 = 0.6; 4, 14 = 6.8; 5, 6 = 10.3; 6, 8ax = 1.5; 8ax, 8eq = 14.0; 8ax, 9eq = 5.4; 8ax, 9ax = 13.6; 8eq, 9eq = 1.6; 8eq, 9ax = 5.0; 9ax, 9eq = 13.6.

(90 MHz, CDCl₃, TMS as internal standard) revealed the presence of signals centred at δ : 4.73 (1H, dd, J = 12 and 0.5 Hz, collapsing to a d on irradiation at δ 2.63 and to an s(br) on irradiation at δ 1.30) consistent with the sequence HC-CH-CH-O-; 2.63 (1H, m, partially obscured by other signals); 1.24 (3H, d, d = 6 Hz collapsing to a s on irradiation at δ 2.63) accounting for O=C-CH(CH₃)-CH-CHOR and 1.24 (3H, s) for a tertiary methyl group.

All the above data showed a close similarity between compounds 1 and 2 with the lack in the latter of the -CH=CH- system.

Detailed examination of the ¹H NMR spectra obtained with a 500 MHz instrument (CDCl₃) led to the conclusion that the remaining hydroxyl group (see IR spectral data)

Table 2. ¹³C NMR chemical shifts of compound 2 (67.88 MHz, CDCl₃, TMS as internal standard)

С	δ	С	δ
1*	37.11 t†	8*	19.91 t
2*	19.91 t	9*	39.76 t
3	211.19 s	10	35.45 s
4	45.55 d	11	135.34 s
5	56.22 d	12	170.11 s
6	80.69 d	14*	13.60 q
7	133.63 s	15*	16.85 q

*Assignments made by comparison with available reference compounds [1, 5]. †SFORD multiplicity.

had to be on the lactone ring as in the case of compound 1. The fact that in these spectra the signals were resolved into an approximately first-order pattern permitted selective decoupling experiments which confirmed, besides the complete assignment of every proton (see Table 1), the C-11 position of the hydroxyl group. Moreover, the particular coupling constants found for H-4, H-5 and H-6 led to the stereochemistry shown at C-4, C-5 and C-6.

The trans-A/B ring junction was proved by the ORD spectrum (0.002 M MeOH, 28°) which showed a positive Cotton effect ($[\alpha]_{320} + 450^{\circ}, [\alpha]_{240} + 980^{\circ}$) as reported in the literature [3, 4] for similar model compounds. Finally, the 13 C NMR (CDCl₃) spectrum of 2 (see

Finally, the ¹³C NMR (CDCl₃) spectrum of 2 (see Table 2) showed 14 carbon atoms possessing complexity and chemical shifts in perfect agreement with the proposed structure.

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